

ACNFP advice on the safety of 2'-fucosyllactose (2'-FL) as a novel food for use in food and food supplements

Reference Number RP1476

Regulated Product Dossier Assessment

Assessment finalised: 10th of June 2024

Summary

An application was submitted to the Food Standards Agency (FSA) and Food Standards Scotland (FSS) in March 2022 from Kyowa Hakko Bio Company Ltd, Japan ("the applicant") for the authorisation of 2'-fucosyllactose (2'-FL) as a novel food.

The novel food is intended to be used as a source of human identical milk oligosaccharide, 2'-FL.

2'-FL is manufactured by microbial fermentation using a genetically modified strain of *Escherichia coli* W, and then refined to yield the purified powder.

This new application is seeking to use the novel food within the following food categories: dairy products and analogues, bakery wares, table-top sweeteners, foods for special groups, beverages, and food supplements. Food supplements are not intended to be used if other foods with added 2'-FL or breast milk are consumed the same day.

The intended uses and use levels are the same as those that have already been authorised for 2'-FL produced by fermentation with genetically modified strains of *E. coli* BL21 (DE3), *E. coli* K-12 DH1, and *Corynebacterium glutamicum* ATCC 13032 (assimilated Commission Implementing Regulation (EU) 2017/2201,

assimilated Commission Implementing Regulation (EU) 2019/388 and assimilated Commission Implementing Regulation (EU) 2023/859, respectively). However, this application is also seeking to use 2'-FL as a food supplement for infants, which is not currently authorised.

To support the FSA and FSS in their evaluation of the application, the Advisory Committee on Novel Foods and Processes (ACNFP) were asked to review the safety dossier and supplementary information provided by the applicant. The Committee did not consider any potential health benefits or claims arising from consuming the food, as the focus of the novel food assessment is to ensure the food is safe, and not putting consumers at a nutritional disadvantage.

The Committee concluded that the applicant had provided sufficient information to assure the novel food was safe under the proposed conditions of use. The anticipated intake levels and the proposed use in food and food supplements was not considered to be nutritionally disadvantageous.

1. Introduction

1. The ACNFP assessed the food safety risks of the novel food and its production under the proposed uses, in line with Article 7 of assimilated Commission Implementing Regulation (EU) 2017/2469. The regulatory framework and the technical guidance put in place by the European Food Safety Agency (EFSA) for full novel food applications is retained as the basis and structure for the assessment (EFSA NDA Panel, 2021).

2. In March 2022, Kyowa Hakko Bio Company Ltd, Japan ("the applicant") submitted a full novel food application for the authorisation of 2'-fucosyllactose (2'-FL). The novel food is a water soluble white to off-white powder composed of $\geq 82.0\%$ w/w dry matter (DM) of 2'-FL, which is manufactured by microbial fermentation using a genetically modified strain of *Escherichia coli* W. The 2'-FL is intended to be used as a source of human identical milk oligosaccharide.

3. Following the review by the ACNFP in September 2023, further information was requested from the applicant concerning the identity, the production process, the compositional information, the stability, the nutritional information, and allergenicity information on the novel food, in order to address information gaps in the initial dossier. The final advice from the Committee was agreed at the 165th meeting, allowing the FSA and FSS to complete the risk assessment.

4. The Committee advice document (CAD) outlines the conclusions of the ACNFP on the safety of the novel food.

2. Assessment

2.1 Identity of the novel food

5. The novel food is a white to off-white powder which is mainly composed of 2'-FL ($\geq 82.0\%$ w/w DM). Other saccharides are present in smaller quantities: D-lactose ($\leq 5\%$ w/w), fucosylgalactose ($\leq 3\%$ w/w DM), difucosyllactose ($\leq 3\%$ w/w DM), L-fucose ($\leq 1\%$ w/w DM), D-glucose and D-galactose ($\leq 1\%$ w/w DM) and a small fraction of other carbohydrates (sum of other carbohydrates $\leq 8.0\%$ w/w DM).

6. 2'-FL is a trisaccharide consisting of L-fucose, D-glucose and D-galactose, (see figure 1). This is identical to the structure of 2'-FL in human breast milk.

Figure 1: The structural formula of 2'-FL

7. 2'-FL is classified as a purified chemical substance and characterised by the following information:

IUPAC name (2*S*,3*S*,4*R*,5*S*,6*S*)-2-[(2*S*,3*R*,4*S*,5*R*,6*R*)-4,5-dihydroxy-6-(hydroxymethyl)-2-[(2*R*,3*S*,4*R*,5*R*)-4,5,6-trihydroxy-2-(hydroxymethyl)oxan-3-yl]oxyoxan-3-yl]oxy-6-methyloxane-3,4,5-triol

CAS number 41263-94-9

Molecular weight 488.44 g/mol

Molecular formula C₁₈H₃₂O₁₅

8. The structure of 2'-FL in the novel food was confirmed using liquid chromatography – tandem mass spectrometry (LC-MS/MS) and nuclear magnetic resonance (NMR) spectroscopy: ¹H-NMR, ¹³C-NMR, and two-dimensional (2D) NMR studies including COSY (correlation spectroscopy), TOCSY (total correlation spectroscopy), HETCOR (heteronuclear correlation) and HMBC (heteronuclear multiple bond correlation) methodologies.

9. 2D-NMR studies using high purity standards of 2'-FL and 3-FL, a conformational isomer of 2'-FL, demonstrated that the L-fucose is linked to D-galactose by α -(1''-2') bonds. This provides unequivocal evidence on the structure of 2'-FL.

10. High-performance liquid chromatography-pulsed amperometric detection (HPLC-PAD) was used to characterise 2'-FL in nine batches of novel food – six batches manufactured using soy peptone in the fermentation media and three batches manufactured without soy peptone in the fermentation media.

2.2 Production Process

11. The production microorganism, *E. coli* SGR5, used to manufacture the novel food is a genetically modified derivative of *E. coli* W (Waksman's strain) that functions as a processing aid as defined in Article 3(2)(b) of assimilated Regulation (EC) No.1333/2008 on food additives. A novel food produced by a GMO does not fall under the remit of the GMO legislation, assimilated Regulation (EC) No 1829/2003 or assimilated Regulation (EC) No 1830/2003, when the production microorganism is removed during the manufacturing process and therefore no recombinant DNA remains. This has been confirmed in the compositional analysis as detailed below.

12. The novel food is classified as category 1 under the EFSA GMO guidance: chemically defined purified compounds and their mixtures in which both genetically modified microorganisms (GMMs) and newly introduced genes have been removed, under EFSA guidance, which categorises GMMs and their products for risk assessment purposes (EFSA GMO Panel, 2011), which the FSA have retained for the purposes of technical review.

13. Although *E. coli* is not considered to be suitable for qualified presumption of safety (QPS) status (EFSA BIOHAZ Panel, 2023), *E. coli* W is widely used for biotechnological applications. Genomic analysis confirms that the genes required for pathogenicity are missing key components or they have been mutationally inactivated (Archer et al., 2011). Furthermore, *E. coli* W is considered to be a safe and non-pathogenic microorganism because this does not cause disease in healthy adult humans or colonise the human gut (Bauer et al., 2008; NIH, 2019). On the basis of this information, the new production strain organism does not introduce any new risks that need to be evaluated and managed.

14. The absence of bacteria from the *Enterobacteriaceae* family (ISO 21528-1:2017) and residual bacterial DNA (LOQ = 4 µg/kg) confirms the genetically modified *E. coli* SRG5 is not present in the novel food.

15. The first stage of the production process involves the conversion of D-lactose and D-glucose to 2'-FL by the adapted cellular metabolism of the production microorganism (The commercial production process does not use soy peptone).

Glucose acts as an exclusive energy and carbon source, and lactose as a substrate for the biosynthesis of 2'-FL. The 2'-FL is released from the *E. coli* SGR5 into the fermentation broth. The production microorganism, *E. coli* SGR5, is removed from the culture medium by microfiltration at the end of the fermentation process.

16. The second stage of the production process involves a series of purification and isolation steps (filtration, ion exchange, concentration and spray drying) to obtain the final purified novel food in powder form.
17. Information on the acceptance criteria for the raw materials and processing aids was provided. The purity criteria for the reagents used in the manufacture of 2'-FL are listed in Table 1.

Table 1: Purity criteria for reagents used to synthesize the novel food

Raw material	Purity
Glucose (powdered)	$\geq 89\%$ (dextrose equivalent)
D-Lactose	Optical rotation $[\alpha]_{20D}$ $+54.4$ to $+55.9^\circ$

18. The novel food is produced in line with Hazard Analysis and Critical Control Point (HACCP) principles. The manufacturing facility is FSSC (Food Safety System Certification) 22000 certified.
19. Bacterial contamination of the novel food is controlled by monitoring the purification steps and the membrane filtration step prior to spray drying. To prevent bacterial growth, the water content in the finished product is monitored and specified at $\leq 9\%$ w/w. Endotoxin levels in the novel food are controlled by ultrafiltration and compliance with the specified level at ≤ 10 EU/mg.
20. The production process has characterised the potential hazards and the corresponding control measures are appropriate.

2.3 Compositional information

21. Results from six independent batches of novel food manufactured using fermentation media containing soy peptone and three independent batches of novel food manufactured using fermentation media without soy peptone

(commercial production process) were provided. These results demonstrated that the novel food is appropriately characterised (Table 2).

Table 2. Compositional Analysis of the novel food produced with and without soy peptone in fermentation media

Test Parameter	Batch 1	Batch 2	Batch 3	Batch 4	Batch 5	Batch 6	Batch 7	Batch 8	Batch 9
2'-FL (% w/w DM)	92	92	92	91	96	94	94	93	91
D-lactose (% w/w DM)	3.1	2.7	2.3	2.4	2.1	2.3	1.1	1.1	1.1
L-fucose (% w/w DM)	≤ 0.05	0.1	0.1	0.1	0.1	0.1	0.05	0.05	0.05
D-glucose and D-galactose (% w/w DM)	0.2	0.1	0.1	0.2	≤ 0.05	0.1	0.3	0.3	0.3
Fucosylgalactose (% w/w DM)	0.8	0.5	0.4	0.9	0.1	0.8	1.0	1.0	1.0
Difucosyllactose (% w/w DM)	0.5	1.4	0.9	1.0	1.1	1.0	0.4	0.4	0.4
Sum of other carbohydrates (% w/w DM)	3.15	3.1	4.1	4.4	0.45	1.6	3.1	4	6.1
Water (% w/w DM)	5.0	3.9	3.9	2.7	2.8	2.3	4.7	5.3	4.5

Test Parameter	Batch 1	Batch 2	Batch 3	Batch 4	Batch 5	Batch 6	Batch 7	Batch 8	Batch 9
Protein (mg/kg)	≤ 100	-----	-----	-----	-----	-----	≤ 100	≤ 100	≤ 100
Ash (% w/w DM)	0.2	0.1	0.1	≤ 0.03	0.1	0.1	0.1	0.2	0.1
pH (5% solution, 25°C)	6.3	6.4	6.2	5.7	6.1	6.2	6.1	6.0	6.2
Arsenic (mg/kg)	≤ 0.05	≤ 0.05	≤ 0.05	≤ 0.05	≤ 0.05	≤ 0.05	0.01	0.01	0.01
Cadmium (mg/kg)	≤ 0.05	≤ 0.05	≤ 0.05	≤ 0.05	≤ 0.05	≤ 0.05	0.01	0.01	0.01
Lead (mg/kg)	≤ 0.05	≤ 0.05	≤ 0.05	≤ 0.05	≤ 0.05	≤ 0.05	0.02	0.02	0.02
Mercury (mg/kg)	≤ 0.05	≤ 0.05	≤ 0.05	≤ 0.05	≤ 0.05	≤ 0.05	0.004	0.004	0.004
Aflatoxin M1 (μ g/kg)	≤ 0.02	-----	-----	-----	-----	-----	≤ 0.02	≤ 0.02	≤ 0.02
Aerobic plate count (CFU/g)	40	10	10	10	10	10	10	10	10
Yeast and mould (CFU/g)	100	100	100	100	100	100	10	10	10
Enterobacteriaceae (in 10g)	ND	ND	ND	ND	ND	ND	ND	ND	ND
<i>Salmonella</i> (in 25g)	ND	-----	-----	ND	ND	ND	ND	ND	ND

Test Parameter	Batch 1	Batch 2	Batch 3	Batch 4	Batch 5	Batch 6	Batch 7	Batch 8	Batch 9
<i>Cronobacter</i> spp. (in 10g)	ND	ND	ND	ND	ND	ND	ND	ND	ND
<i>Listeria</i> <i>monocytogenes</i> (in 25g)	ND	ND	ND	ND	ND	ND	ND	ND	ND
Presumptive <i>Bacillus cereus</i> (CFU/g)	10	10	10	10	10	10	10	10	10
Endotoxins (EU/mg)	0.0092	0.0005	0.0080	0.0028	0.0005	0.0003	0.0002	0.0002	0.0002

Batches 1 – 6 manufactured with soy peptone in fermentation media; Batches 7 – 9 manufactured without soy peptone in fermentation media

FL = fucosyllactose; DM = dry matter; ND = not detected; ---- = not determined; CFU = colony forming units; EU/mg = endotoxin units/mg

Limit of quantification (LOQ) = 0.05% w/w for D-lactose, L-fucose, D-glucose and D-galactose, fucosylgalactose and difucosyllactose. LOQ = 0.03% w/w for ash

Sum of other carbohydrates (% w/w DM) = 100 – (2'-FL + D-lactose, L-fucose, D-glucose and D-galactose, fucosylgalactose, difucosyllactose + ash).

Protein test limit = 100 mg/kg; LOQ for heavy metals = 0.05 mg/kg (batches 1 – 6) and LOQ for As and Cd = 0.01 mg/kg; Pb = 0.02 mg/kg; Hg = 0.01 mg/kg (batches 7 – 9); endotoxins = 0.0001563 EU/mg (rounded up to 0.0002). Aerobic plate count LOD = 10 CFU/g; in accordance with ISO 4833-1:2013, the presence of 1 to 3 colonies should be reported as 40 CFU/g; yeast and mould LOD = 100 CFU/g (surface plating) for batches 1 – 6 and LOD = 10 CFU/g (in-depth plating) for batches 7 – 9; LOD for Presumptive *Bacillus cereus* = 10 CFU/g.

22. The composition data demonstrates that the novel food is consistently the same when the manufactured with or without soya peptone in the fermentation media.

23. Lactose is the most abundant saccharide component in human milk. The breakdown products of lactose, glucose and galactose, are normal constituents of human milk. L-fucose is one of the building blocks of 2'-FL. DFL is a member of the fucosylated oligosaccharides that constitute around 70% of the total HMO fraction in human milk (Bode, 2012). Fucosylgalactose is a natural component produced by the action of the fucosyltransferase enzyme (EFSA NDA Panel, 2023).

24. An assessment was conducted in accordance with the EFSA Guidance on technical requirements for regulated food and feed product applications to establish the presence of small particles including nanoparticles in the novel food (EFSA, 2021). A solubility in water test was and the results confirmed that the novel food exceeded the decision criteria for solubility (> 33.3 g/L) as described in the guidance. Therefore, further evaluation for the presence of nanoparticles in the novel food is not required.

25. Certification was provided to demonstrate that the contract laboratories were accredited to perform these analytical studies. Where in-house analysis was utilised, full methodology and supporting validation documentation was provided.

26. The data presented indicate the novel food and any hazards present were appropriately characterised.

2.4 Stability

27. Results from an ongoing 36-month real-time stability study ($25 \pm 2^{\circ}\text{C}$; $60 \pm 5\%$ Relative Humidity) on a single batch of novel food produced with soy peptone in the fermentation media was provided. Data covering 30 months was reported for 2'-FL. Testing for, carbohydrate, water content, physicochemical parameters, and water activity was undertaken at the 6-, 12- and 24-month time-points. These endpoints in addition to microbiology quality were assessed at 30 months. No significant changes were observed, and microbial parameters were below the limits of detection.

28. A 6-month accelerated stability study ($40 \pm 2^{\circ}\text{C}$; $75 \pm 5\%$ Relative Humidity) was conducted on five batches of novel food produced with soy peptone in the fermentation media. Data for the following parameters was provided: 2'-FL, carbohydrate, water content, and physicochemical parameters and water activity.

Microbiological data was provided for two batches. No significant changes were observed.

29. No stability data was provided for the novel food in different food matrices.

30. The presence of soy peptone in the fermentation media is not expected to have a significant impact on the composition of the novel food. On this basis, the data provided supports the stability of the novel food, manufactured with or without soy peptone in the fermentation media, for up to 30 months.

2.5 Specification

31. The specification parameters for the novel food (Table 3) were assessed using internationally recognised methods or are otherwise determined using internally developed and validated methods.

Table 3: Specification for the novel food

Parameter	Specification	Method
2'-FL	≥ 82.0 % w/w DM	HPLC-CAD (in-house)
D-lactose	≤ 5.0 % w/w DM	HPLC-CAD (in-house)
L-fucose	≤ 1.0 % w/w DM	HPLC-CAD (in-house)
D-glucose and D-galactose	≤ 1.0 % w/w DM	HPLC-CAD (in-house)
Fucosylgalactose	≤ 3.0 % w/w DM	HPLC-CAD (in-house)
Difucosyllactose	≤ 3 % w/w DM	HPLC-CAD (in-house)
Sum of other carbohydrates	≤ 8.0 % w/w DM	Calculation
Water	≤ 9.0 % w/w DM	JP 2.48

Parameter	Specification	Method
Protein	≤ 0.01 % w/w DM	Bradford assay
Ash	≤ 0.5 % w/w DM	JP 2.44
pH (5% solution, 25°C)	4.5 – 8.5	JP 2.54
Arsenic	≤ 0.2 mg/kg	USP 233> or AOAC method
Cadmium	≤ 0.1 mg/kg	USP 233> or AOAC method
Lead	≤ 0.02 mg/kg	USP 233> or AOAC method
Mercury	≤ 0.1 mg/kg	USP 233> or EPA method
Aflatoxin M1	≤ 0.025 µg/kg	AOAC 2000.08
Aerobic plate count)	≤ 1,000 CFU/g	ISO 4833-1:2013
Yeast and mould	≤ 100 CFU/g	ISO 21527-2:2008
Enterobacteriaceae	Absent in 10g	ISO 21528-1:2017
<i>Salmonella</i> spp.	Absent in 25g	ISO 6579-1:2017
<i>Cronobacter</i> spp.	Absent in 10g	ISO 22964:2017
<i>Listeria monocytogenes</i>	Absent in 25g	ISO 11290-1:2017
Presumptive <i>Bacillus cereus</i>	≤ 50 CFU/g	ISO 7932:2004

Parameter	Specification	Method
Endotoxins	≤ 10 EU/mg	JP 4.01

DM = dry matter; CFU = colony forming units; HPLC-PAD = high-performance liquid chromatography – pulsed amperometric detection; JP = Japanese Pharmacopoeia; USP = United States Pharmacopoeia; EPA = Environmental Protection Agency; ICP-MS = Inductively coupled plasma – mass spectrometry; ICP-OES = Inductively coupled plasma – optical emission spectroscopy; AOAC = Association of Official Analytical Chemists; ISO = International Organisation for Standardisation

D-Glucose and D-galactose peaks on the HPLC-PAD chromatograms overlap.

Sum of other carbohydrates (% w/w DM) = 100 – (2'-FL + D-lactose, L-fucose, D-glucose and D-galactose, fucosylgalactose, difucosyllactose + ash).

Heavy metals (As, Cd and Pb) – AOAC (2019) 999.10 and 2011.14; Hg – EPA (2007) 7473

32. The information provided is sufficient for the specification of the novel food, and appropriately characterises the novel food seeking authorisation.

2.6 History of Use

33. There is no history of use for the novel food in the UK.

34. 2'-FL, which is a major constituent of the novel food, has been authorised in the EU (assimilated Commission Implementing Regulation (EU) 2017/2201, assimilated Commission Implementing Regulation (EU) 2019/388 and assimilated Commission Implementing Regulation (EU) 2023/859) for use in a range of foods and food supplements. The authorised 2'-FL is produced by fermentation using a genetically modified strains of *E. coli* BL21 (DE3), *E. coli* K-12 DH1, and *Corynebacterium glutamicum* ATCC 13032, respectively.

35. Human breast milk contains a family of structurally related oligosaccharides, known as human milk oligosaccharides (HMOs), as the third largest solid components (Kunz and Rudloff, 1993; Bode, 2012; Newburg, 2013). The concentrations of HMOs in human colostrum are 20 to 25 g/L, whereas in mature human milk, the concentrations are 5 to 20 g/L (Thurl *et al.*, 2010; Bode, 2012;

Urashima *et al.*, 2018).

36. 2'-FL belongs to the subfraction of fucosylated HMOs (Thurl *et al.*, 2010; Rijnierse *et al.*, 2011; Bode, 2012), and is one of the most predominant HMOs in human milk, with a mean of mean concentration of 2.28 g/L and a maximum mean concentration of 4.28 g/L (Soyyilmaz *et al.* (2021). Higher intakes may occur as even higher concentrations of 2'-FL have been reported in human milk (up to 4.78 g/L – Thurl *et al.*, 2017; 5.57 g/L – Austin *et al.*, 2019 and 5.85 g/L Samuel *et al.*, 2019).

37. Using the reported levels of 2'-FL in human breast milk from Soyylmaz *et al.* (2021) and considering the average and high daily intake of breast milk (800 mL and 1,200 mL, respectively) for infants from 0 to 6 months (EFSA NDA Panel, 2013), the daily intake levels of 2'-FL from human milk for a 6.7 kg body weight infant (EFSA SC, 2012) are shown in Table 4.

Table 4: Estimated intakes of 2'-FL from 800ml and 1200ml of Breast milk for 6.7kg Infant ^a using data from Soyylmaz *et al.* (2021)

	Mean intake levels (mg/kg BW/day)	95th percentile intake levels (mg/kg BW/day)
Estimated highest intake for 800ml milk *	272	511
Estimated highest intake for 1200ml milk *	408	767

* EFSA NDA Panel, 2013

38. The history of use does not indicate any further areas for evaluation.

2.7 Proposed Use and Anticipated Intake

39. The target population is the general population.

40. The intended uses and use levels of the 2'-FL from the novel food are listed in Table 5. These food categories and intended use levels are the same as those in

the List of Novel Foods (assimilated Commission Implementing Regulation (EU) 2017/2470). However, the use of 2'-FL as a food supplement for infants is not currently authorised.

41. The anticipated intake for 2'-FL in children up to the age of 16 weeks is estimated to be 312 mg/kg body weight/day for a 6.7 kg infant. This value was calculated from the use of 2'-FL in infant formula (1.2 g/L) at a high consumption level of 260 ml/kg body weight/day, as established by the EFSA Scientific Committee (EFSA SC, 2017). This value does not exceed the estimated high daily intake of 2'-FL in breast-fed infants per kg/BW (see Table 4).

Table 5: Food Categories and Use Levels for 2'-FL from the novel food

Food Category Name	Proposed Maximum Use Level
Unflavoured pasteurised and unflavoured sterilised (including UHT) milk products	1.2 g/L
	1.2 g/L beverages
Unflavoured fermented milk-based products	19.2 g/kg for products other than beverages
	1.2 g/L beverages
Flavoured fermented milk-based products including heat-treated products	19.2 g/kg for products other than beverages
	1.2 g/L beverages
Dairy analogues, including beverage whiteners	12.0 g/kg for products other than beverages
	400 g/kg for whitener
Fine bakery wares. Cereal bars only	12.0 g/kg

Food Category Name	Proposed Maximum Use Level
Table-top sweetener	200 g/kg
Infant formula as defined in assimilated Regulation (EU) No 609/2013	1.2 g/L in the final product ready for use, marketed as such or reconstituted as instructed by the manufacturer
Follow-on formula as defined in assimilated Regulation (EU) No 609/2013	1.2 g/L in the final product ready for use, marketed as such or reconstituted as instructed by the manufacturer
Processed cereal-based food and baby food for infants and young children as defined in assimilated Regulation (EU) No 609/2013	1.2 g/L in the final product ready for use, marketed as such or reconstituted as instructed by the manufacturer 12.0 g/kg for products other than beverages
Milk-based drinks and similar products intended for young children	1.2 g/L (beverages) in the final product ready for use, marketed as such or reconstituted as instructed by the manufacturer
Foods for special medical purposes as defined in assimilated Regulation (EU) No 609/2013	In accordance with the particular nutritional requirements of the persons for whom the products are intended

Food Category Name	Proposed Maximum Use Level
Total diet replacement for weight control as defined in assimilated Regulation (EU) No 609/2013	4.8 g/L beverages (equivalent to 1.2 g/meal based on a standard 250 g/meal replacement beverage) 40 g/kg for products other than beverages (equivalent to 1.20 g/meal based on a standard 30 g meal replacement bar)
Bread and pasta products bearing statements on the absence or reduced presence of gluten in accordance with the requirements of assimilated Commission Implementing Regulation (EU) No 828/2014	60 g/kg
Flavoured drinks	1.2 g/L
Coffee, tea (excluding black tea), herbal and fruit infusions, chicory; tea, herbal and fruit infusions and chicory extracts; tea, plant, fruit and cereal preparations for infusions, as well as mixes and instant mixes of these products	9.6 g/L – the maximum level refers to the products ready to use
Food supplements *	3.0 g/day for general population; 1.2 g/day for infants and young children

* Food Supplements as defined in the Food Supplements (England) Regulations 2003, the Food Supplements (Wales) Regulations 2003 and the Food Supplements (Scotland) Regulations 2003.

42. An intake assessment for 2'-FL was conducted in support of the original application using the UK National Diet and Nutrition Survey (NDNS) for years

2008–2010 (EFSA NDA Panel, 2015a). For this reason, an updated intake assessment was conducted using the EFSA Dietary Exposure (DietEx) tool, which is based on individual data from the EFSA Comprehensive European Food Consumption Database.

43. The estimated mean and high-level intakes of 2'-FL from the proposed conditions of use for each sub-population are presented in Table 6.

Table 6: Estimated daily intake for 2'-FL from the current authorised uses in the List of Novel Foods (assimilated Commission Implementing Regulation (EU) 2017/2470).

Population Group	Mean Intakes of 2'-FL (mg/kg BW/day)	95th percentile intakes of 2'-FL (mg/kg BW/day)
Infants (≤ 11 months)	51 – 438	132 – 1,377
Young children (12 to 35 months)	116 – 398	292 – 869
Other children (3 to 9 years)	45 – 198	140 – 605
Adolescents (10 to 17 years)	14 – 81	52 – 233
Adults (18 to 64 years)	36 – 153	95 – 337

44. The highest estimated intake for 2'-FL from human breast milk consumption is 767 mg/kg BW/day in infants (Table 4). The estimated 95th percentile intakes in the infant and young children sub-populations are 1,377 mg/kg BW/day and 869 mg/kg BW/day.

45. The use level for 2'-FL in food supplements for infants is 1.2 g/day. In young children (1.2 g/day) and the general population (3.0 g/day), the use of 2'-FL in food supplements is already authorised (EFSA NDA Panel, 2015a, b).

46. In infants weighing 5kg (EFSA Scientific Committee, 2012), the maximum intake of 2'-FL in food supplements for infants would be 240 mg/kg BW/day. On this basis, consumption of 2'-FL in food supplements is not expected to exceed the levels found in human breast milk (Table 4).

47. Food supplements containing the novel food are not intended to be used if other foods containing 2'-FL, including breast milk, are consumed on the same day.

2.8 Absorption, Distribution, Metabolism and Excretion (ADME)

48. No ADME studies were conducted on the novel food.

49. 2'-FL does not undergo any significant digestion by human enzymes in the upper gastrointestinal tract and only small amounts are expected to be absorbed. HMOs are fermented in the colon by intestinal microbiota with a fraction excreted unchanged in the faeces and a small fraction found in the urine (EFSA NDA Panel, 2022). There is no information which indicates that 2'-FL in the novel food differs from the 2'-FL in human breast milk (EFSA NDA Panel, 2023).

50. The ADME of human milk oligosaccharides are well understood and the information does not indicate any further areas of concern.

2.9 Nutritional information

51. The novel food is mainly composed of the oligosaccharide, 2'-FL, which is structurally identical to the naturally occurring counterpart in human breast milk.

52. Consumption of the novel food at the proposed use levels is not expected to be nutritionally disadvantageous for consumers.

2.10 Toxicological information

53. Toxicological studies were performed to support the safety assessment of 2'-FL. The respective study reports are unpublished and claimed as proprietary data. They were reviewed by the ACNFP and considered essential in the assessment of the safety of the novel food.

2.10.1 Genotoxicity

54. *In vitro* genotoxicity testing of 2'-FL was conducted under Good Laboratory Practice (GLP) conditions and according to the OECD guidelines: *in vitro* bacterial reverse mutation test (OECD TG 471) and *in vivo* mammalian erythrocyte micronucleus test (OECD TG 474). This is not the approach recommended by the UK Committee on Mutagenicity or in the guidance on the preparation and submission of an application for authorisation of a novel food in the context of assimilated Regulation (EU) 2015/2283.

55. The Scientific opinion on genotoxicity testing strategies applicable to food and feed safety assessment (EFSA, 2011) recommends two *in vitro* tests as the first step in testing for genotoxicity: *in vitro* bacterial reverse mutation test (OECD TG 471) and *in vitro* mammalian cell micronucleus test (OECD TG 487). This approach addresses the key endpoints for adequately assessing genotoxicity with the minimum number of tests and avoiding unnecessary animal tests.

56. The *in vivo* mammalian erythrocyte micronucleus test (OECD TG 474) assesses both structural and numerical chromosomal aberrations and is an appropriate follow-up test for *in vitro* clastogens and aneugens.

57. The *in vitro* bacterial reverse mutation test (Ogama, 2019a [unpublished]) demonstrated that 2'-FL is non-mutagenic at concentrations up to 5,000 µg 2'-FL/plate, in the absence or presence of metabolic activation.

58. The *in vivo* mammalian micronucleus test (Ogama, 2019b [unpublished]) reported that 2'-FL is non-clastogenic and non-aneugenic. However, no evidence was provided to demonstrate that the bone marrow in mice had been exposed to 2'-FL. Therefore, an *in vitro* micronucleus test was conducted.

59. The *in vitro* micronucleus test (Kikuchi, 2022 [unpublished]) demonstrated that 2'-FL is non-clastogenic and non-aneugenic in the absence or presence of metabolic activation up to the highest concentration of 2,000 µg 2'-FL/ml.

60. The results from these *in vitro* and *in vivo* studies support the conclusion that 2'-FL is not genotoxic.

2.10.2 Sub-chronic toxicity

61. A Repeated Dose 90-day oral gavage study in rodents (Tsuboi, 2020 [unpublished]) was conducted under GLP conditions according to OECD TG 408 guidelines as recommended by the Guidance on the preparation and submission of an application for authorisation of a novel food in the context of assimilated Regulation (EU) 2015/2283. The aim of the study was to identify any adverse

effects following the consumption of the novel food.

62. In this 90-day oral gavage study, each group consisted of 10 female and 10 male Crl:CD(SD) rats which were dosed with 0 (control – vehicle only [water]), 500, 1,000 or 2,000 mg/kg BW/day of 2'-FL by oral gavage.

63. No deaths, test item-related clinical abnormalities, or differences in food consumption and bodyweight between test groups were reported. There were no statistically significant dose dependent changes in clinical chemistry, haematology, serum hormone levels, urinalysis, or organ weights.

64. An increase in the incidence of ocular changes were reported. However, the applicant has confirmed these were within the historical control range for the facility and therefore not of concern.

65. No dose related abnormalities were noted during the necropsy or histopathological evaluation. Therefore, the no observable adverse effect level (NOAEL) for 2'-FL was considered to be the highest dose tested of 2,000 mg/kg BW/day..

2.10.3 Human studies

66. No human clinical trials were conducted with the novel food.

2.11 Allergenicity

67. The protein content of the novel food is reported as 0.01% w/w.

68. Absence of bacteria from the *Enterobacteriaceae* family (ISO 21528-1:2017) confirmed that the genetically modified *E. coli* SRG5 is not present in the novel food.

69. The potential allergenicity of the introduced proteins expressed in *E. coli* W was assessed using the National Institute of Health Sciences (Japan) Allergen Database for Food Safety and conducted in line with FAO (2001) guidelines. None of the proteins was predicted to be an allergen.

70. Two ELISA (enzyme linked immunosorbent assay) tests were conducted on the novel food manufactured using soy peptone to detect the presence of milk proteins (LOQ = 1 µg/g). The results confirmed that milk proteins were effectively removed during the purification process and were not present in the finished powder.

71. Five batches of the novel food manufactured using soy peptone were also tested for the presence of soybean protein. The results from the ELISA test (LOQ = 1 µg/g) confirmed that soybean protein was effectively removed from all batches of the novel food during the purification process and was not present in the finished powder.

72. The likelihood of allergenic reactions to the novel food is expected to be low under the proposed conditions of use.

3. Discussion

73. The novel food is a white to off-white powder which is mainly composed of the human identical milk trisaccharide, 2'-FL ($\geq 90.0\%$ w/w DM), as well as other saccharides in smaller quantities.

74. The novel food is manufactured by microbial fermentation using a genetically modified strain of *Escherichia coli* W and then refined to yield the purified powder.

75. 2'-FL is intended to be used in dairy products and analogues, bakery wares, table-top sweeteners, foods for special groups, beverages, and food supplements. The general population are identified as the target population of the novel food.

76. Analysis confirms that the 2'-FL is structurally identical to the 2'-FL found naturally in human milk. Exposure to 2'-FL relates solely to breastfeeding infants as there is no recognised history of use for this milk oligosaccharide as an ingredient in foods or food supplements.

77. In the Repeated Dose 90-day oral gavage study in rodents, the NOAEL for 2'-FL was 2,000 mg/kg BW/day, the highest dose tested. When this NOAEL is compared with the highest estimated exposure in each population category, the margins of exposure range from 1.5 to 8.6. Given that the 2'-FL in the novel food is equivalent to 2'-FL found in human breast milk, these margins of exposure are acceptable with respect to the highest estimated daily intakes in the intended population.

78. For most sub-populations, the estimated daily intake of 2'-FL is not expected to exceed the highest estimated intake level of 767 mg/kg BW/day for 2'-FL in breastfed infants. In the infant and young children sub-populations, the highest estimated 95th percentile intake of 2'-FL is reported as 1,377 mg/kg BW/day and 869 mg/kg BW/day, which both exceed this value. However, given the wide range of estimated intakes for 2'-FL, the limited absorption of HMOs, and the absence of

toxicological effects reported in the *in vivo* 90-day oral gavage study, this is not considered to be a concern.

79. The use level of 2'-FL in food supplements (1.2 g/day for infants and young children, and 3.0 g/day for all other sub-populations) is not expected to exceed the highest intake level of 2'-FL in breastfed infants on a body weight basis. Food supplements are not intended to be used by the general population if other foods containing the novel food, including breast milk or other foods for infants and young children, are consumed on the same day.

4. Conclusion

80. The ACNFP have undertaken the assessment of the novel food, which is composed mainly of 2'-FL, and concluded that the novel food is safe under the proposed conditions of use and does not pose a safety risk to human health. The anticipated intake level and the proposed use in food and food supplements was not considered to be nutritionally disadvantageous.

81. The advice was based on the information in the novel food dossier submitted by the applicant plus the supplementary information and could not have been reached without the following data claimed as proprietary by the applicant:

- annexes to the dossier which relate to the identity of the novel food, the production process, composition, stability, and the solubility of the novel food
- *in vitro* bacterial reverse mutation test (Ogama, 2019a [unpublished]); *in vitro* micronucleus test (Kikuchi, 2022 [unpublished]); 90-day repeat dose oral gavage study with the novel food (Tsuboi, 2020 [unpublished])

The members of the ACNFP during the course of the assessment who were;

Dr Camilla Alexander White, Dr Anton Alldrick, Dr Kimon Andreas Karatzas, Ms Alison Austin, Professor George Bassel, Dr Mark Berry, Dr Christine Bosch, Professor Dimitris Charalampopoulos, Dr Catharina Edwards, Professor Susan Fairweather-Tait, Professor Paul Frazer, Dr Hamid Ghoddusi, Professor Andy Greenfield, Professor Wendy Harwood, Professor Huw D. Jones, Dr Ray Kemp, Dr Elizabeth Lund, Professor Harry J. McArdle, Mrs Rebecca McKenzie, Professor Clare Mills, Dr Antonio Peña-Fernández, Dr Lesley Stanley, Professor Hans Verhagen, Dr Maureen Wakefield, and Professor Bruce Whitelaw.

References

Archer CT, Kim JF, Jeong H, Park JH, Vickers CE, Lee SY and Nielsen LK, 2011. The genome sequence of E. coli W (ATCC 9637): comparative genome analysis and an improved genome-scale reconstruction of E. coli. BMC Genomics, 12, 20.

<https://doi.org/10.1186/1471-2164-12-9>

Austin S, De Castro CA, Sprenger N, Binia A, Affolter M, Garcia-Rodenas CL, Beauport L, Tolsa JF and Fischer Fumeaux CJ, 2019. Human milk oligosaccharides in the milk of mothers delivering term versus preterm infants. Nutrients, 11, 1282.

<https://doi.org/10.3390/nu11061282>

Bauer AP, Ludwig W and Schleifer KH, 2008. A novel DNA microarray design for accurate and straightforward identification of Escherichia coli safety and laboratory strains. Systematic and Applied Microbiology, 31, 50–61.

<https://doi.org/10.1016/j.syapm.2008.01.001>

Bode L, 2012. Human milk oligosaccharides: every baby needs a sugar mama. Glycobiology, 22(9), 1147-1162. <https://doi.org/10.1093/glycob/cws074>

EFSA BIOHAZ Panel (EFSA Panel on Biological Hazards), 2023. Scientific Opinion on the update of the list of qualified presumption of safety (QPS) recommended microorganisms intentionally added to food or feed as notified to EFSA. EFSA Journal 2023;21(1):7747, 23 pp. <https://doi.org/10.2903/j.efsa.2023.7747>

FSA GMO Panel (EFSA Panel on Genetically Modified Organisms), 2011. Scientific Opinion on Guidance on the risk assessment of genetically modified microorganisms and their products intended for food and feed use (Question no EFSA-Q-2009-00521, adopted: 25 May 2009). EFSA Journal, 9(6):2193 [54 pp.].

<https://doi.org/10.2903/j.efsa.2011.2193>

EFSA NDA Panel (EFSA Panel on Dietetic Products, Nutrition and Allergies), 2013. Scientific Opinion on nutrient requirements and dietary intakes of infants and young children in the European Union. EFSA Journal, 11(10):3408 [103 pp.].

<https://doi.org/10.2903/j.efsa.2013.3408>

EFSA NDA Panel (EFSA Panel on Dietetic Products, Nutrition and Allergies), 2015a. Scientific opinion on the safety of 2'-O-fucosyllactose as a novel food ingredient pursuant to Regulation (EC) No 258/97. EFSA Journal 13 (7):4184, 32 pp.

<https://doi.org/10.2903/j.efsa.2015.4184>

EFSA NDA Panel (EFSA Panel on Dietetic Products, Nutrition and Allergies), 2015b. Statement on the safety of lacto-N-neotetraose and 2'-O-fucosyllactose as novel food ingredients in food supplements for children. EFSA Journal 13(11):4299, 11 pp. <https://doi.org/10.2903/j.efsa.2015.4299>

FSA NDA Panel (EFSA Panel on Nutrition, Novel Foods and Food Allergens), 2019. Scientific opinion on the dietary reference values for sodium. EFSA Journal 2019; 17(9): 5778, [191 pp]. <https://doi.org/10.2903/j.efsa.2019.5778>

EFSA NDA Panel (Panel on Dietetic Products, Nutrition and Allergies), 2021. Guidance on the preparation and submission of an application for authorisation of a novel food in the context of Regulation (EU) 2015/2283 (Revision 1). Published 26 March. <https://doi.org/10.2903/j.efsa.2011.2170>

EFSA NDA Panel (EFSA Panel on Nutrition, Novel Foods and Food Allergens), 2022. Scientific Opinion on the safety of 20-fucosyllactose (2'-FL) produced by a derivative strain (APC199) of *Corynebacterium glutamicum* ATCC 13032 as a Novel Food pursuant to Regulation (EU) 2015/2283. EFSA Journal 2022;20(12):7647, 21 pp. <https://doi.org/10.2903/j.efsa.2022.7647>

EFSA NDA Panel (EFSA Panel on Nutrition, Novel Foods and Food Allergens), 2023. Safety of 2'-fucosyllactose (2'-FL) produced by a derivative strain (*Escherichia coli* SGR5) of *E. coli* W (ATCC 9637) as a Novel Food pursuant to Regulation (EU) 2015/2283. EFSA Journal, 21 (11), 1-26. <https://doi.org/10.2903/j.efsa.2023.8333>

EFSA, 2011. Scientific Opinion on genotoxicity testing strategies applicable to food and feed safety assessment. EFSA Journal 9(9):2379 [69 pp.]

<https://doi.org/10.2903/j.efsa.2011.2379>

EFSA, 2012. Scientific Committee Guidance on selected default values to be used by the EFSA Scientific Committee, Scientific Panels and Units in the absence of actual measured data. EFSA Journal 10(3):2579. [32 pp.]

<https://doi.org/10.2903/j.efsa.2012.2579>

EFSA Scientific Committee, 2017. Guidance on the risk assessment of substances present in food intended for infants below 16 weeks of age (Question no EFSA-Q-2016-00489, adopted: 26 April 2017). EFSA Journal, 15(5):4849 [58 pp.].

<https://doi.org/10.2903/j.efsa.2017.4849>

EFSA, 2021. Scientific Committee on Guidance on technical requirements for regulated food and feed product applications to establish the presence of small particles including nanoparticles. EFSA Journal 19(8):6769 [48 pp.]

<https://doi.org/10.2903/j.efsa.2021.6769>

FAO/WHO, 2001. Evaluation of allergenicity of genetically modified foods. Report of a Joint FAO/WHO Expert Consultation on Allergenicity of Food Derived from Biotechnology, 22-25 January 2001. Food and Agriculture organisation of the United Nations (FAO), Rome, Italy. https://cdn.who.int/media/docs/default-source/documents/publications/evaluation-of-allergenicity.pdf?sfvrsn=9528a2fb_2&download=true (Accessed March 2024).

Kikuchi M, 2022 [unpublished]. Dated 26 September 2022. Yoshimi Laboratories, Drug Safety Testing Center Co., Ltd, Saitama 355-0166, Japan. Study Title: Micronucleus Study of 2'-Fucosyllactose (2FL Na) in Cultured Mammalian Cells (Study number: CG220002)

Kunz C and Rudloff S, 1993. Biological functions of oligosaccharides in human milk. Acta Paediatrica, 82(12), 903-912.

<https://doi.org/10.1111/j.1651-2227.1993.tb12597.x>

Newburg DS, 2013. Glycobiology of human milk. Biochemistry (Moscow), 78, 771-785 (Russian edition "Biokhimiya, 778, 990-1007").

<https://doi.org/10.1134/s0006297913070092>

NIH (National Institutes of Health), 2019. NIH guidelines for research involving recombinant or synthetic nucleic acid molecules. National Institutes of Health (NIH), Office of Science Policy, Bethesda, MD Available online:

<https://osp.od.nih.gov/biotechnology/nih-guidelines/> (accessed on 1 March 2024)

OECD, 1997. Bacterial reverse mutation test. In OECD guidelines for the testing of chemicals. OECD guideline No 471 (updated & adopted: 21 July 1997). Paris, France: Organisation for Economic Co-operation and Development (OECD).

<https://doi.org/10.1787/9789264071247-en>

OECD, 1998. OECD principles of good laboratory practice. Series on principles of good laboratory practice and compliance monitoring, No. 1 (ENV/MC/CHEM(98) 17). Paris, France: Organisation for Economic Co-operation and Development (OECD), Environment Directorate, Chemicals Group and Management Committee.

<https://doi.org/10.1787/9789264078536-en>

OECD, 2016a. Mammalian *in vivo* micronucleus test. In OECD guidelines for the testing of chemicals. OECD guideline No 474 (updated & adopted: 29 July 2016). Paris, France: Organisation for Economic Co-operation and Development (OECD).

<https://doi.org/10.1787/9789264264762-en>

OECD, 2016b. *In vitro* mammalian cell micronucleus test. In OECD guidelines for the testing of chemicals. OECD guideline No 487 (updated & adopted: 29 July 2016). Paris, France: Organisation for Economic Co-operation and Development (OECD).

<https://doi.org/10.1787/9789264264861-en>

OECD, 2018. Repeated dose 90-day oral toxicity study in rodents. In OECD guidelines for the testing of chemicals. OECD guideline No 408 (updated and adopted 27 June 2018). Paris, France: Organisation for Economic Co-operation and Development (OECD). <https://doi.org/10.1787/9789264070707-en>

Ogama Y, 2019a [unpublished]. Dated 18 October 2019. Yoshimi Laboratories, Drug Safety Testing Center Co., Ltd, Saitama 355-0166, Japan. Study Title: A BACTERIAL REVERSE MUTATION TEST OF 2'-Fucosyllactose (Study number: AG190035).

Ogamai Y, 2019b [unpublished]. Dated 13 December 2019. Yoshimi Laboratories, Drug Safety Testing Center Co., Ltd, Saitama 355-0166, Japan. Study Title: Micronucleus Study of 2'-Fucosyllactose in Bone-marrow Cells of Mice (Study number: 200058)

Rijnierse A, Jeurink PV, van Esch BCAM, Garssen J and Knippels LMJ, 2011. Food-derived oligosaccharides exhibit pharmaceutical properties. European Journal of Pharmacology, 668, S117–S123. <https://doi.org/10.1016/j.ejphar.2011.07.009>

Samuel TM, Binia A, de Castro CA, Thakkar SK, Billeaud C, Agosti M, Al-Jashi I, Costeira MJ, Marchini G, Martinez-Costa C, Picaud JC, Stiris T, Stoicescu SM, Vanpe_e M, Domell€of M, Austin S and Sprengel N, 2019. Impact of maternal characteristics on human milk oligosaccharide composition over the first 4 months of lactation in a cohort of healthy European mothers. Scientific Reports, 9, 11767.

<https://doi.org/10.1038/s41598-019-48337-4>

Soyyılmaz B, Miks MH, Reohrig CH, Matwiejuk M, Meszaros-Matwiejuk A and Vignæs LK, 2021. The mean of milk: a review of human milk oligosaccharide concentrations throughout lactation. *Nutrients*, 13, 2737.

<https://doi.org/10.3390/nu13082737>

Thurl S, Munzert M, Henker J, Boehm G, Muller-Werner B, Jelinek J and Stahl B, 2010. Variation of human milk oligosaccharides in relation to milk groups and lactational periods. *British Journal of Nutrition*, 104, 1261–1271.

<https://doi.org/10.1016/j.bbi.2015.06.025>

Thurl S, Munzert M, Boehm G, Matthews C and Stahl B, 2017. Systematic review of the concentrations of oligosaccharides in human milk. *Nutrition Reviews*, 75(11), 920-933. <https://doi.org/10.1093/nutrit/nux044>

Tsuboi M, 2020 [unpublished]. Dated 27 January 2020. Yoshimi Laboratories, Drug Safety Testing Center Co., Ltd, Saitama 355-0166, Japan. Study Title: Ninety-Day Repeated Oral Dose Toxicity Study of 2'-fucosyllactose in Rats (Study number: 100602RG).

Urashima T, Yamaguchi E, Ohshima T, Fukuda K and Saito T, 2018. Chemical structures of oligosaccharides in milk of the raccoon (*Procyon lotor*). *Glycoconjugate Journal*, 35, 275–286. <https://doi.org/10.1007/s10719-018-9821-z>